# **Project Descriptions**

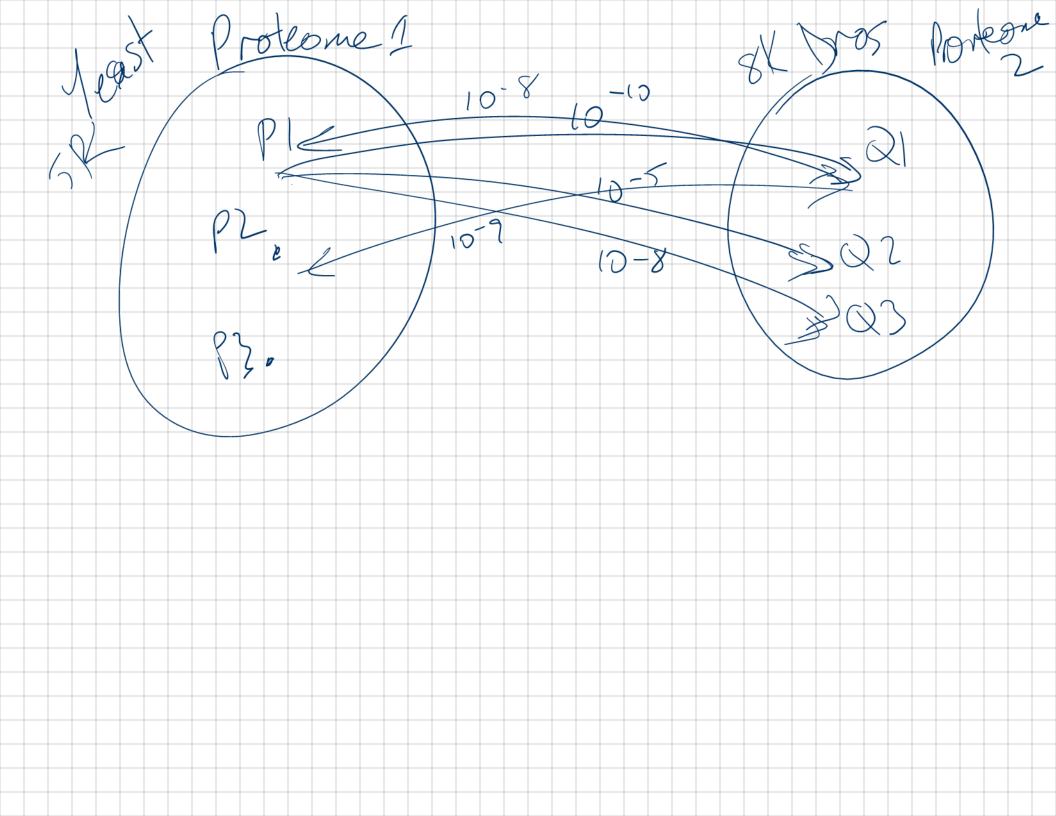
#### BINF5240 Lecture 18

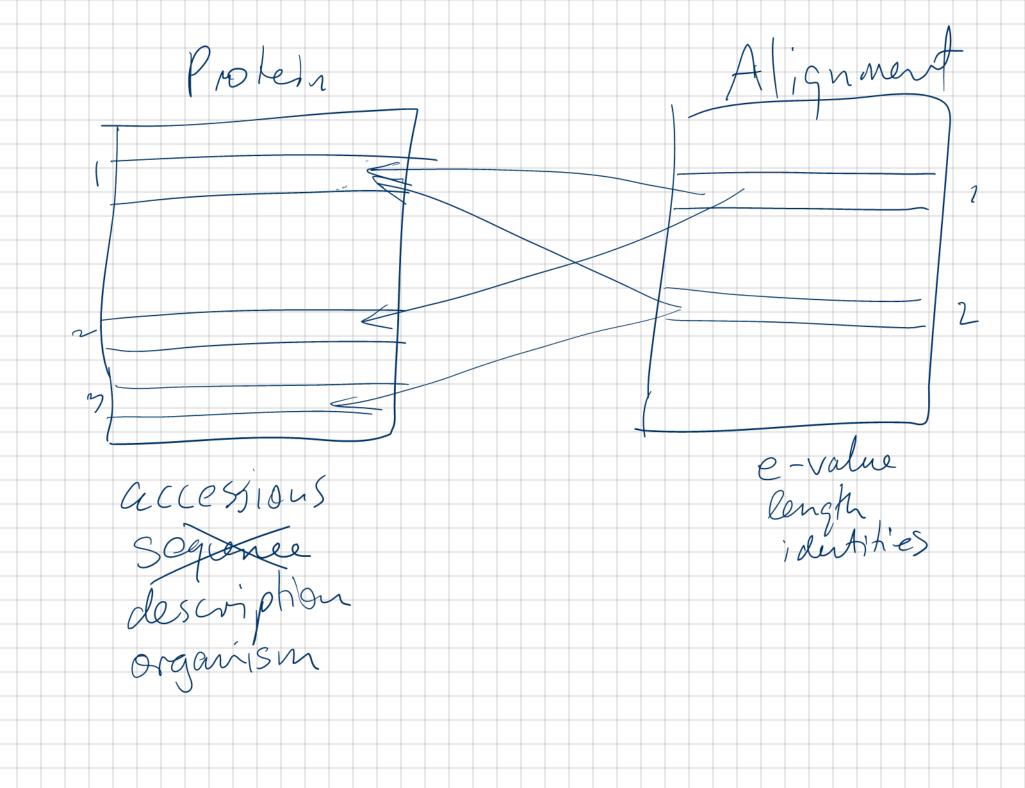
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## **Class Project: Blast Database**

- 1. Write a program that computes all pairwise blast alignments for two species' proteomes and stores the alignments in a relational database.
- 2. Write a program that retrieves the blast alignment for two proteins (specified by their accessions) from the relational database.
- Write a program that finds pairs of orthologous proteins that are mutually best hits in the species' proteomes.







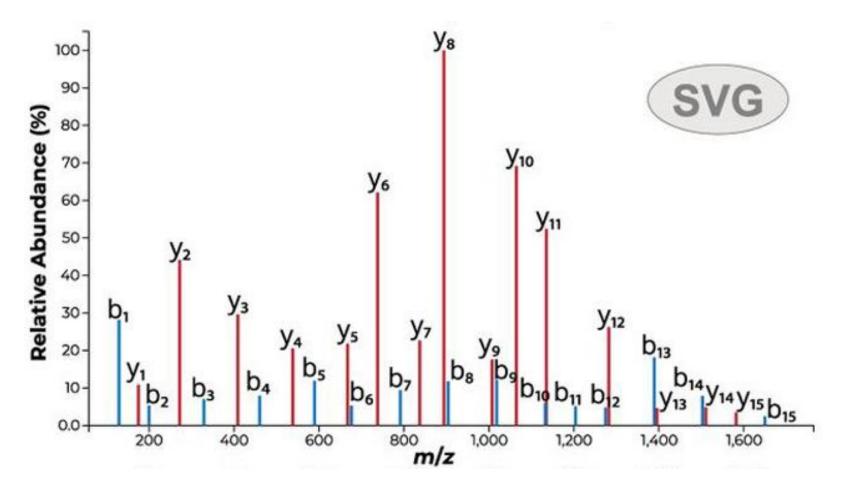
## **Class Project: MS/MS Viewer**

- Write a program to display peptide fragmentation spectra from an mzXML file.
  - The program will take an mzXML file, a scan number, and a peptide sequence as input.
  - The peptide's b-ion and y-ion m/z values should be computed, and peaks matching these m/z values annotated with appropriate labels.
  - The output figure/plot should aid the user in determining whether or not the peptide is a good match to the spectrum.

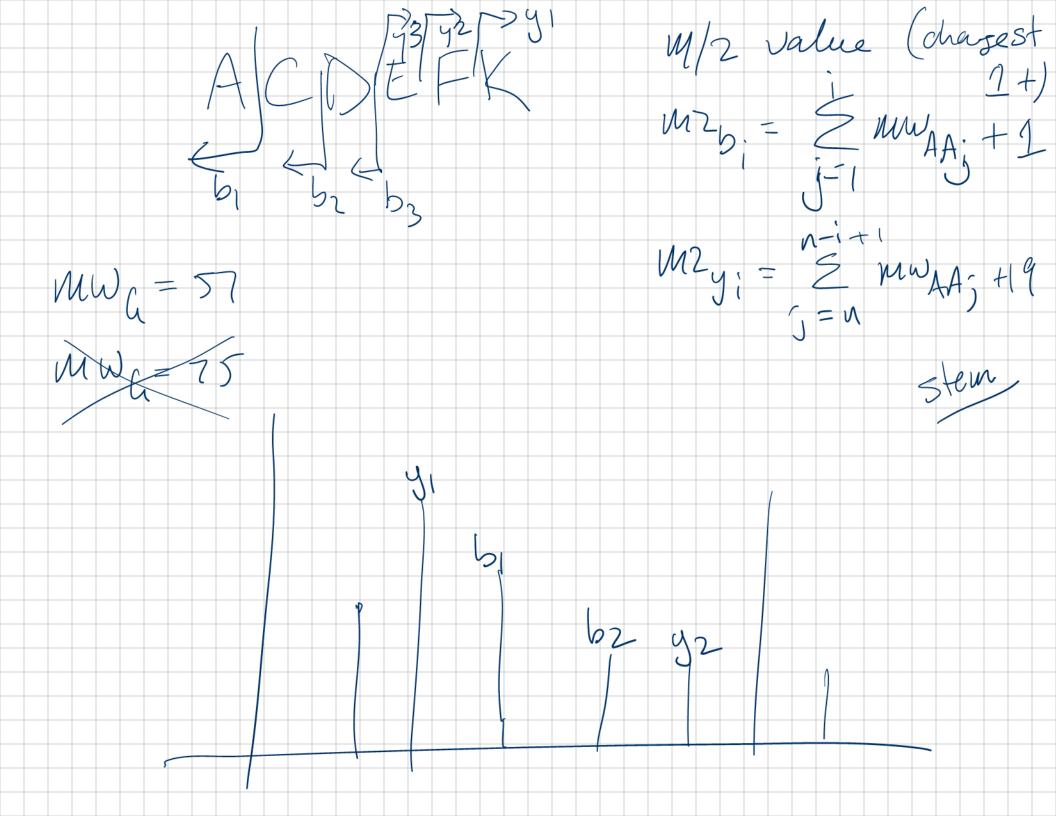


# Example of annotated spectrum





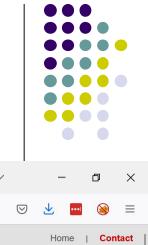
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# **Class Project: Protein Digest**

- Write a simple web-server application using TurboGears to carry out an *in silico* enzymatic digest of a user-provided protein sequence.
  - Users should be able to specify min and max length, min and max molecular weight, # of missed cleavages, and specific enzyme.
  - Output should be a table of peptides, with their length, molecular weight, # of missed cleavages, and amino-acids to left and right of each peptide in the protein sequence.





### **Example: PeptideMass**

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Expasy <sup>3</sup>	PeptideMass		Home	Con	tact

#### **PeptideMass**

**PeptideMass** [references] cleaves a protein sequence from the UniProt Knowledgebase (Swiss-Prot and TrEMBL) or a user-entered protein sequence with a chosen enzyme, and computes the masses of the generated peptides. The tool also returns theoretical isoelectric point and mass values for the protein of interest. If desired, PeptideMass can return the mass of peptides known to carry post-translational modifications, and can highlight peptides whose masses may be affected by database conflicts, polymorphisms or splice variants.

#### Instructions are available.

Enter a UniProtKB protein identifier, ID (e.g. ALBU\_HUMAN), or accession number, AC (e.g. P04406), or an amino acid sequence (e.g. 'SELVEGVIV'; you may specify post-translational modifications, but PLEASE read this document first!):

		/
	/	//.

Reset the fields. Perform the cleavage of the protein.

The peptide masses are

with cysteines treated with: nothing (in reduced form)  $\checkmark$ 

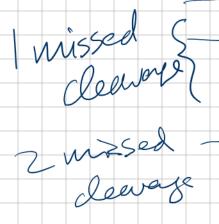
T with convlomide adducts

https://web.expasy.org/peptide\_mass/



### **Example: PeptideMass**

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$\leftarrow$	ightarrow G	0	https://web. <b>expasy.org</b> /cgi-bin/peptide_mass/peptide-mass.pl		$\bigtriangledown$	⊻	••••  (	8	≡
	1149.5759 25-34	1	DAHKSEVAHR						
	698.3580 29-34	0	SEVAHR						
	973.5214 29-36	1	SEVAHRFK						
	1226.6051 35-44	1	FKDLGEENFK						
	951.4418 37-44	0	DLGEENFK						
	3365.6874 37-65	1	DLGEENFKALVLIAFAQYLQ QCPFEDHVK						
	2433.2635 45-65	0	ALVLIAFAQYLQQCPFEDHV K						
	3563.8606 45-75	1	ALVLIAFAQYLQQCPFEDHV KLVNEVTEFAK						
	1149.6150 66-75	0	LVNEVTEFAK						
	2515.1326 66-88	1	LVNEVTEFAKTCVADESAEN CDK						
	1384.5355 76-88	0	TCVADESAENCDK						
	2383.0540 76-97	1	TCVADESAENCDKSLHTLFG DK						
	1017.5363 89-97	0	SLHTLFGDK						
	1875.0156 89-105	1	SLHTLFGDKLCTVATLR						
	876.4971 98-105	0	LCTVATLR						
	2177.9698 98-117	1	LCTVATLRETYGEMADCCAK						
	1320.4905 106-117	0	ETYGEMADCCAK						
	1959.7881 106-122	1	ETYGEMADCCAKQEPER						
	658.3155 118-122	0	QEPER						
	1657.7751 118-130	1	QEPERNECFLQHK						
	1018.4775 123-130	0	NECFLQHK						
	1939.9079 123-138	1	NECFLQHKDDNPNLPR						
	940.4483 131-138	0	DDNPNLPR						
	0544 0700 404 400								



MW(pep) = sum(amino-acids) + 19 # [M+H]